

International Journal of Environment and Climate Change

Volume 13, Issue 7, Page 10-23, 2023; Article no.IJECC.99288 ISSN: 2581-8627

(Past name: British Journal of Environment & Climate Change, Past ISSN: 2231-4784)

GxE analysis to Identify the Stable High-yielding Rice Lines among a Set of Selected Germplasm Panel

Padmashree R. ^{a,b}, Vishal Reddy ^{a,c}, Kalyani M. Barbadikar ^{a*}, Honnappa M. ^{a,b}, Nakul D. Magar ^a, Divya Balakrishnan ^a, C. Gireesh ^a, Anantha M. Siddaiah ^a, Jyothi Badri ^a, R. Lokesha ^c, Y. M. Ramesha ^d, P. Senguttuvel ^a, J. R. Diwan ^e, Maganti Sheshu Madhav ^{a,f} and R. M. Sundaram ^a

^a ICAR-Indian Institute Rice Research (IIRR), Hyderabad-500030, India.

^b Department of Genetics and Plant Breeding, UAS, Raichur-584104, India.

^c Keladi Shivappa Nayaka University of Agricultural and Horticultural Sciences,
Shivamogga-577412, India.

^d Department of Agronomy, ARS Dhadesugur, UAS, Raichur-584167, India.

^e Department of Genetics and Plant Breeding, ARS Mugad, UAS, Dharwad-580007, India.

^f Central Tobacco Research Institute, Raiahmundry-533106, India.

Authors' contributions

This work was carried out in collaboration among all authors. The study was conceived and planned by author KMB. Phenotyping of the panel was executed by authors PR, HM and NDM. Phenotypic and genotypic data analysis were done by authors PR, VR, DB, KMB, CG, MSM, AMS and YMR. The critical editing was done by authors KMB, PS, MSM, RL, JRD and RMS. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJECC/2023/v13i71849

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here:

https://www.sdiarticle5.com/review-history/99288

Original Research Article

Received: 18/02/2023 Accepted: 21/04/2023 Published: 26/04/2023

ABSTRACT

Rice lines need to be grown and evaluated for yield under different agro-ecological locations to identify stable and high-yielding lines for deployment in breeding programs. With this aim, a set of rice germplasm was evaluated for GxE in four different environments (E1-Dadesuguru-Wet 2020, E2-ICAR-IIRR-Dry 2019, E3-ICAR-IIRR-Wet 2020, E4-ICAR-IIRR-Dry 2020). The experimental trial was laid out in a randomized complete block (RCB) design with three replications at each location for 118 rice lines. Data on yield per plant was analyzed using the Additive Main Effect and Multiplicative Interaction (AMMI) and Genotype, and Genotype x Environment Interaction (GGE) models. The combined analysis of variance (ANOVA) manifested significant variations for tested genotypes, locations, years, genotype x year, and genotype x location interactions revealing the influence of environmental factors on yield traits. All four environments showed discrimination power, whereas E2 and E3 were found as the representative environment as they fall near the Average-Environment axis (AEA). The AMMI biplot PC1 contributed 79.20% variability and PC2 contributed 15.18% variability. From the GGE biplot analysis, the rice lines Phouren, JBB-631-1, and JBB-1325 were found to be the best and most stable. The rice lines Phouren, PUP-229, and TI-112 were stable in the first sub-group Dhadesugur-Wet 2020 (E1). The rice lines Langphou, and NPK-45 were stable in the second sub-group ICAR-IIRR-Wet 2020 (E3). Environment ICAR-IIRR-Dry 2019 (E2) was the third subgroup and the rice lines Moirangphou-Yenthik and TI-3 topped for the same. The ICAR IIRR-Dry 2020 (E4) environment formed the fourth subgroup where Phouren-Amubi, TI-128 and JBB-1325 topped the season. In conclusion, this study revealed that G x E interactions are significant for yield variation, and its AMMI and biplots analysis are efficient tools for visualizing the response of genotypes to different locations.

Keywords: Rice; AMMI; G x E interaction; stability.

1. INTRODUCTION

Grain yield is a complex quantitative trait, with high environmental interaction. The selection of lines for breeding needs to be made based on the performance under multiple environments, especially for rice which is grown under a wide range of ecosystems. It is essential to carry out selection based on yield stability evaluation rather than average performance in multiple environment conditions [1]. Selection genotypes for stability and adaptability required prior to the recommendation. Stability is the suitability of a variety over a wide range of environments while adaptability is the better survival of a genotype over any specific environment [2]. There are several methods to study stability and genotype x environment interactions of traits through conventional analysis. Different models were proposed on stability variance. covalence, regression coefficient analysis or principal component analysis (PCA) [3]. Eberhart and Russell [4], Perkins and Jinks [5], Freeman and Perkins [6], Shukla [7], Kang, [8]. However, the additive main effects and multiplicative interaction (AMMI) model and the genotype main effects and genotype x environment interaction effects (GGE) model are more popular methods. This method is followed to quantify the genotypeenvironment interaction through PCA and

graphical representation and has been widely applied in multi-environment cultivar trials [9] Crossa et al. [10] Gauch and Zobel, [11]. The breeding protocol must quantify genotype, environment, and their interaction factors to obtain successful breeding results of yield and related traits in rice [2].

An evaluation of phenotypic traits (P) assessed upon multiple environments (E) will be different from an assessment of genotypic traits (G), so $P = G + E + (G \times E)$. Due to the huge impact that GxE interactions have on phenotype, it is imperative to conduct stability analyses to assess genotype performance in various environments and make the process easier for plant breeders. Therefore, the estimation of the GxE interaction. on the other hand, has continuously an intriguing factor in plant breeding. Biplots are presently genotypeto evaluate broadly utilized environment interaction (GEI) since the GxE impacts can be envisioned in a single chart, making genotypes and their intelligence with situations less demanding to compare [12]. Yan et al. [13] proposed a modern GEI examination procedure based on biplots, comparative to the AMMI method, that has the advantage of dissecting down the combined impact of genotype (G) and $G \times E(G + GE)$ by central component examination, as restricted to the initial AMMI investigation.

The essential ANOVA depicts fundamental impacts and determines whether genotype x environment may be a critical source of variability, but it does not give knowledge into the variation caused by genotypes or environment. Besides, PCA contains no sources of variations for genotype or environment and does not viably dissect interactions. AMMI analysis [11] improves the likelihood of effective selection and has been used to assess genotype x environment interaction with greater precision in various crops [2]. The GGE biplot is regularly utilized to discover the GEI variation generated in the experimental design. It clearly simplifies the testing locations for large environments. The goal of this study was to figure out how studied four environments affect yield in specific genotypes and to find sites that are best suited for higher production. As a result, it offers the optimum genotype for rice growers in the region, as well as for the country's familiar agro-ecologies.

2. MATERIALS AND METHODS

The experimental material consists of 118 rice lines including North-Eastern landraces, aromatic genotypes, advanced breeding lines, soft rice lines, mutants of BPT-5204 and Nagina 22 (N22), wild introgression lines, *tropical japonica* accessions, *Oryza glaberrima* accessions and popular varieties (Table 1) [14].

2.1 Field Experiments

The experiments were conducted for four seasons at two different locations to evaluate the rice lines (E1-Dadesuguru-wet 2020, E2-ICAR-IIRR-Dry 2019, E3-ICAR -IIRR-wet 2020, E4-ICAR -IIRR-Dry 2020). The experimental trial was laid out in a randomized complete block (RCB) design with three replications at each location for 118 rice lines. At each replication, 25-30 days-old seedlings were transplanted into 13 rows with a spacing of 20 x15cm, and the recommended package of practices for rice production in the region was followed [14]. Data on yield traits were analyzed using the Additive Main Effect and Multiplicative Interaction (AMMI), Genotype and Genotype x Environment Interaction (GGE) as mentioned by Balakrishnan et al. [2].

2.2 Additive Main and Multiplicative Interaction (AMMI) Analysis

Using the statistical program R studio, the GxE interaction of 118 lines in four environments was

assessed using the AMMI model proposed by Gauch and Zobel in [11]. After fitting an ANOVA model with the main effects of genotype and (without environment interaction). standardized residuals were used to fit a principal component analysis (PCA). experimental error as well as the influence of the GxE interaction is included in these residuals. The equation was: 1. $Y_{i_j} = \mu + G_i + E_j + \sum \lambda_k \, \alpha_{i_k} \, \gamma_{j_k} \, + e_{i_j}$ where Y_{i_j} is the observed mean yield of the ith genotype in the jth environment, m is the general mean, G_i represents the effects of genotypes and environments for the k^{th} axis, α_{i_k} is the eigenvector of the i^{th} genotype for the k^{th} axis, γ_{j_k} represents the eigenvector of the j^{th} environment for the k^{th} axis, n is the number of principal components in the model, $e_{i,j}$ is the average of the corresponding random errors. The contribution of IPCA1 and IPCA2 sums of squares to the interaction sum of squares was used to compute it for each genotype [15]. The formula for calculating the AMMI stability value is as follows: 2. $ASV = sqrt((\frac{SS_{pc1}}{SS_{pc2}} \times PC1_i)^2 + (PC2_i)^2)$ where, SS_{pc1} and SS_{pc2} are the sums of squares of pc1 and pc2 respectively, and the score and pc2 score are the scores of the genotype in those particular PCAs.

2.3 GGE Bi-plot Analysis

In order to interpret GEI patterns, we used the GGE bi-plot methodology, which is a combination of AMMI bi-plot and GGE concepts. An average-environment coordination (AEC) view of biplots based on environment-focused scaling was used to interpret mean genotype performance *vs* their adaptability patterns from the polygon view of genotype-environment interaction biplots of symmetrical scaling [16].

3. RESULTS AND DISCUSSION

The yield per plant exhibited a considerable contribution from main factors (genotypes and environments) and interaction effects, according to the AMMI model. A significant mean sum of squares attributable to lines suggested that there were genotypic differences, and the relevance of the environment explains why environmental effects change across different sites and test locations. Furthermore, genotype-environment interaction results indicate that lines behave differently in different seasons (Table 1). Based on the mean yield performance and stability of

the rice lines, three North-Eastern landraces Phouren, Longphou, Phouren-Amubi and Moirangphou-Yenthink, two advanced breeding lines derived from *indica /tropical japonica* crosses JBB-631-1 and JBB-1325, a Near isogenic line, PUP 229, two mutants of BPT-5204 TI-112 and TI-3 and wild introgression line NPK-45 were stable.

The AMMI plot demonstrated that the rice lines Phouren, JBB-631-1, and JBB-1325 were stable for grain yield per plant, which is in agreement with the study of Sharifi et al. [17] that reported seven stable rice lines for grain yield stability in three consecutive years. Similarly, Ashwini et al. [18] used the AMMI plot to analyze conventional and enhanced rice varieties in five districts of Karnataka. In the case of yield, it was observed that genotypic mean fluctuations were far higher than environmental mean variations. Dewi et al. [19] also found that when rice genotypes were examined in different growing seasons, the mean sum of squares due to the genotype's main effect was high for grain yield.

The multiplicative variance of the treatment sum of squares due to the GE interaction was significant and was further partitioned into the interaction principal component axis (IPCA). The IPCA I and IPCA II scores, explained 79.20 per cent and 15.18 per cent of the interaction. The total GEI for the trait grain yield per hectare was captured by these two PCA axes, which accounted for 94.38% of the total GEI. Sharifi et al. [17] conducted a GxE interaction study in which the proportion of IPC1 (49%) to the interaction sum of squares was far greater than that of IPC2 (24%).

3.1 Stability Parameters

The rice lines Phouren (26.18 g/plant) had the lowest AMMI stability value (0.68), followed by JBB-631-1 (24.70 g/plant) and JBB-1325 (23.90 g/plant) with 0.75 and 0.89 ASV, respectively. Similarly, according to the Genotypic Selection Index (GSI), Phouren was ranked as the best line as it had a lower GSI value (Table 1).

3.2 GGE Biplots

GGE biplot represents the polygon view of the GGE biplot and it revealed the best-performing genotypes for traits in a single group location [13]. As explained by Oladosu et al. [20] the genotypes lying on the vertex of a polygon with no environmental indicator nearby are poorly performed genotypes, and the genotypes that

are present on the vertex of a polygon where one or more environmental indicators are present are the best-performing genotypes in the relevant environments. From 'Which won where' pattern of GGE biplots for grain yield of chosen varieties revealed that rice lines Phouren, JBB-631-1, and JBB-1325 were found to be best most and stable for grain yield per plant because they were placed at edge of the polygon (Fig. 1). The radiating lines from the biplot origin intersected each of the polygon sides at right angles, dividing the four environments into four sub-groups. The rice lines Phouren, and PUP-229, TI-112 fall in the first sub-group Dhadesugur-Wet 2020 (E1). The rice lines Langphou, NPK-45 the second sub-group in ICAR-IIRR-Wet 2020 (E3). Environment ICAR-IIRR-Dry 2019 (E2) was the third subgroup consisting of the rice lines Moirangphou-Yenthik and TI-3. The ICAR IIRR-Dry 2020 (E4) environment formed the fourth subgroup consisting of Phouren-Amubi, TI-128 and JBB-1325 were found to be stable. This confirms the presence of interaction between genotype and environment for all the traits evaluated. Babu et al. [16] identified genotypes G22 (Edavankudi Pokkali), G17 (Taraori Basmati), G27 (Chittimuthyalu) and G26 (Kalanamak) stable for zinc in polished rice (ZPR) and G8 (Savitri) stable for single plant yield (SPY) across three environments. The findings of our study are the agreement with the report stated by Khan et al. [21] and Mwiinga et al. [22].

The GGE biplot for grain yield per plant, that is descriptiveness vs representativeness (Fig. 2) revealed that the environment Dhadesugur (E1) had the shortest vector, indicating that this environment was unable to discriminate lines, whereas the environments ICAR-IIRR-*Dry* 2019 (E2), ICAR-IIRR-*Wet* 2020 (E3), and ICAR-IIRR-*Dry* 2020 (E4) had longer vectors than Dhadesugur-*Wet* 2020 (E1), indicating that environments were able to discriminate lines. The location, Dhadesugur- *Wet* 2020 (E1) has a limited ability to discriminate, which could be due to environmental or other error factors [16].

The stability pattern across four locations was analyzed using the mean vs stability pattern of the GGE biplot (Fig. 3). The GGE plot facilitates line evaluation based on mean performance and stability across four environments. The biplot graph is formed by the intersection of a vertical AEC abscissa and a horizontal AEC ordinate line. Each line has a single arrowhead that points towards a higher mean performance for the

Table 1. AMMI stability parameters for grain yield/ plant

| SI. No. | Rice lines | Special Characters | IPCA1 | IPCA2 | ASV | Rank ASV | Mean | Rank of | GSI | Rank of |
|---------|-----------------------|----------------------|-------|-------|-------|----------|-------------|---------|-----|---------|
| | | | | | | | Yield/plant | mean | | GSI |
| G1 | KJ-214 | Tropical japonica | 2.41 | -1.99 | 2.95 | 45 | 17.86 | 46 | 91 | 46 |
| G2 | KJ-216 | accessions | -0.62 | 0.51 | 5.91 | 111 | 17.00 | 111 | 222 | 111 |
| G3 | KJ-219 | | -1.53 | -1.42 | 4.42 | 73 | 16.05 | 73 | 146 | 73 |
| G4 | KJ-221 | | -1.84 | 1.74 | 3.49 | 57 | 17.40 | 57 | 114 | 57 |
| G5 | KJ-222 | | 1.41 | -1.19 | 4.68 | 81 | 15.23 | 81 | 162 | 81 |
| G6 | KJ-226 | | -1.72 | -1.71 | 4.17 | 67 | 16.45 | 67 | 134 | 67 |
| G7 | WB-3 (Wangoo-Phou) | North-Eastern | 2.39 | -1.97 | 2.98 | 47 | 17.70 | 47 | 94 | 47 |
| G8 | WB-5 (Phouren) | Landraces | -3.24 | 1.62 | 0.68 | 1 | 26.18 | 1 | 2 | 1 |
| G9 | WB-6 (Chakhao) | | -1.44 | -1.28 | 4.64 | 79 | 15.35 | 79 | 158 | 79 |
| G10 | WB-8 (Moirangphou- | | -3.14 | -2.76 | 1.19 | 8 | 22.10 | 8 | 16 | 8 |
| | Yenthik) | | | | _ | | - | | - | |
| G11 | WB-10 (Langphou) | | 3.16 | -2.98 | 1.05 | 6 | 22.84 | 6 | 12 | 6 |
| G12 | WB-12 (Langphou- | | -0.82 | 0.64 | 5.82 | 106 | 10.52 | 106 | 212 | 106 |
| | Chakao) | | | | | | | | | |
| G13 | WB-14 (Ayangleima) | | -0.29 | -0.39 | 6.86 | 118 | 10.00 | 118 | 236 | 118 |
| G14 | WB-15 (Heimang-Phou) | | -1.46 | -1.29 | 4.57 | 78 | 15.53 | 78 | 156 | 78 |
| G15 | WB-16 (Phouoibi) | | -1.38 | 1.18 | 4.72 | 82 | 15.23 | 82 | 164 | 82 |
| G16 | WB-22 (Moirang-Phou- | | -0.91 | 0.76 | 5.79 | 104 | 11.28 | 104 | 208 | 104 |
| 0.0 | Khokngangbi) | | 0.01 | 00 | 0.70 | | 20 | | 200 | |
| G17 | WB-23 (Kakcheng-Phou) | | -1.15 | 0.93 | 5.35 | 95 | 13.24 | 95 | 190 | 95 |
| G18 | WB-24 (Pat-Phou) | | -2.91 | -2.58 | 2.02 | 22 | 20.10 | 22 | 44 | 22 |
| G19 | WB-26 (Taothabi) | | 2.63 | -2.47 | 2.14 | 26 | 19.73 | 26 | 52 | 26 |
| G20 | WB-27 (Langmanbi) | | -3.12 | -2.66 | 1.56 | 14 | 20.79 | 14 | 28 | 14 |
| G21 | WB-29 (Akut-Phou) | | -2.54 | -2.27 | 2.45 | 34 | 19.16 | 34 | 68 | 34 |
| G22 | WB-30 (MoirangPhou- | | 3.13 | -2.72 | 1.46 | 13 | 20.81 | 13 | 26 | 13 |
| OLL | Angouba) | | 0.10 | | 11.10 | .0 | 20.01 | .0 | | .0 |
| G23 | WB-32 (Keibi-Phou) | | -3.12 | -2.7 | 1.67 | 15 | 20.58 | 15 | 30 | 15 |
| G24 | WB-39 (Phouren-Amubi) | | -3.14 | 2.82 | 1.41 | 10 | 21.66 | 10 | 20 | 10 |
| G25 | GNV-1109 | Advanced breeding | -0.43 | -0.46 | 6.32 | 116 | 12.00 | 116 | 232 | 116 |
| 320 | 0.11 | line | 0.40 | 0.40 | 0.02 | 110 | 12.00 | 110 | 202 | 110 |
| G26 | GNV-1089 | Advanced breeding | -0.61 | 0.58 | 5.96 | 112 | 17.00 | 112 | 224 | 112 |
| 320 | O14V-1003 | line | -0.01 | 0.50 | 5.50 | 114 | 17.00 | 112 | 227 | 112 |
| G27 | RNR-15048 | Popular mega variety | -1.71 | 1.55 | 4.19 | 68 | 16.43 | 68 | 136 | 68 |
| G28 | Pokkali | Cultivated variety | -2.92 | -2.59 | 1.95 | 20 | 20.25 | 20 | 40 | 20 |
| G29 | Siri-1253 | Cultivated variety | -1.19 | -0.97 | 5.03 | 92 | 13.77 | 92 | 184 | 92 |

Padmashree et al.; Int. J. Environ. Clim. Change, vol. 13, no. 7, pp. 10-23, 2023; Article no.IJECC.99288

| SI. No. | Rice lines | Special Characters | IPCA1 | IPCA2 | ASV | Rank ASV | Mean Yield/plant | Rank of mean | GSI | Rank of GSI |
|---------|--------------|-------------------------|-------|-------|------|----------|---------------------|--------------|-----|----------------|
| G30 | GNV-14-96-1 | Advanced breeding lines | -2.5 | 2.18 | 2.6 | 38 | 18.90 | 38 | 76 | 38 |
| G31 | RP-Bio-226 | Cultivated variety | 2.55 | 2.31 | 2.42 | 33 | 19.20 | 33 | 66 | 33 |
| G32 | Tellahamsa | Popular mega variety | -2.57 | 2.35 | 2.38 | 31 | 19.48 | 31 | 62 | 31 |
| G33 | FL-478 | Cultivated variety | 2.53 | 2.23 | 2.53 | 36 | 18.96 | 36 | 72 | 36 |
| G34 | Ratnamudi | Karnataka landrace | -0.78 | 0.64 | 5.88 | 109 | 19.00 | 109 | 218 | 109 |
| G35 | Ratnachudi | | 2.54 | -2.25 | 2.53 | 35 | 19.10 | 35 | 70 | 35 |
| G36 | Tanu | Popular mega varieties | 0.81 | -0.61 | 5.84 | 107 | 10.34 | 107 | 214 | 107 |
| G37 | Rasi | | 2.37 | -1.91 | 3.16 | 49 | 17.61 | 49 | 98 | 49 |
| G38 | Swarna Sub-1 | | 1.77 | -1.64 | 3.62 | 63 | 16.90 | 63 | 126 | 63 |
| G39 | MTU-1010 | | -1.01 | 0.77 | 5.78 | 103 | 11.34 | 103 | 206 | 103 |
| G40 | BPT-5204 | | 1.3 | 1.15 | 4.81 | 85 | 14.76 | 85 | 170 | 85 |
| G41 | Jaya | | -3.11 | 2.69 | 1.78 | 16 | 20.55 | 16 | 32 | 16 |
| G42 | MTU-1001 | | 2.44 | -2.06 | 2.81 | 43 | 17.99 | 43 | 86 | 43 |
| G43 | TI-3 | Mutants of BPT-5204 | -3.14 | 2.74 | 1.36 | 9 | 21.89 | 9 | 18 | 9 |
| G44 | TI-4 | | -0.51 | 0.49 | 6.14 | 114 | 14.00 | 114 | 228 | 114 |
| G45 | TI-8 | | 1.13 | -0.89 | 5.4 | 97 | 13.09 | 97 | 194 | 97 |
| G46 | TI-11 | | -1.09 | 0.83 | 5.51 | 100 | 12.65 | 100 | 200 | 100 |
| G47 | TI-12 | | 1.11 | -0.84 | 5.49 | 99 | 12.69 | 99 | 198 | 99 |
| G48 | TI-15 | | -1.81 | 1.73 | 3.54 | 60 | 17.13 | 60 | 120 | 60 |
| G49 | TI-16 | | 1.54 | -1.44 | 4.36 | 72 | 16.18 | 72 | 144 | 72 |
| G50 | TI-17 | | -1.12 | 0.88 | 5.48 | 98 | 13.08 | 98 | 196 | 98 |
| G51 | TI-18 | | 1.86 | -1.76 | 3.43 | 56 | 17.43 | 56 | 112 | 56 |
| G52 | TI-19 | | -1.37 | 1.16 | 4.76 | 83 | 14.89 | 83 | 166 | 83 |
| G53 | TI-23 | | -0.49 | -0.47 | 6.32 | 115 | 13.00 | 115 | 230 | 115 |
| G54 | TI 24 | | 1.79 | -1.72 | 3.58 | 61 | 17.02 | 61 | 122 | 61 |
| G55 | TI-25 | | -2.49 | 2.17 | 2.63 | 39 | 18.47 | 39 | 78 | 39 |
| G56 | TI 35 | | -1.36 | 1.17 | 4.79 | 84 | 14.78 | 84 | 168 | 84 |
| G57 | TI 36 | | 3.13 | -2.71 | 1.42 | 12 | 20.87 | 12 | 24 | 12 |
| G58 | TI-37 | | -1.27 | 1.13 | 4.88 | 87 | 14.72 | 87 | 174 | 87 |
| G59 | TI-44 | | -1.76 | -1.62 | 3.67 | 64 | 16.65 | 64 | 128 | 64 |
| G60 | TI-87 | | 1.42 | -1.21 | 4.66 | 80 | 15.25 | 80 | 160 | 80 |
| G61 | TI-112 | | 3.17 | 2.18 | 1 | 5 | 23.10 | 5 | 10 | 5 |
| G62 | TI-128 | | -3.13 | -2.78 | 1.42 | 11 | 21.54 | 11 | 22 | 11 |
| G63 | TI-166 | | -1.21 | 1.04 | 5 | 91 | 13.86 | 91 | 182 | 91 |
| G64 | TI-124 | | -1.69 | 1.54 | 4.24 | 69 | 16.38 | 69 | 138 | 69 |

| SI. No. | Rice lines | Special Characters | IPCA1 | IPCA2 | ASV | Rank ASV | Mean Yield/plant | Rank of mean | GSI | Rank of GSI |
|---------|---------------------------------|--|-------|-------|------|----------|---------------------|--------------|-----|----------------|
| G65 | Swarna | Cultivated variety | -1.51 | -1.37 | 4.47 | 75 | 15.71 | 75 | 150 | 75 |
| G66 | Vandana | Cultivated variety | -1.82 | -1.72 | 3.54 | 59 | 17.18 | 59 | 118 | 59 |
| G67 | Wazuhophek | Landrace | -1.52 | -1.43 | 4.43 | 74 | 15.99 | 74 | 148 | 74 |
| G68 | Improved Samba Mahsuri (ISM) | Cultivated variety | -2.38 | -1.96 | 3.15 | 48 | 17.69 | 48 | 96 | 48 |
| G69 | PUP-225 (ISM × VANDANA) | Near isogenic lines | -1.22 | 1.07 | 4.97 | 90 | 14.31 | 90 | 180 | 90 |
| G70 | PUP-229 (MTU1010 × Vandana) | | -3.2 | -3.12 | 0.96 | 4 | 23.50 | 4 | 8 | 4 |
| G71 | PUP-230 (MTU1010 × Vandana) | | -1.9 | -1.8 | 3.38 | 54 | 17.47 | 54 | 108 | 54 |
| G72 | KR-209 (ÍSM × Wazuhophek) | Recombinant inbred lines | -2.56 | 2.32 | 2.4 | 32 | 19.36 | 32 | 64 | 32 |
| G73 | KR-262 (ISM × Wazuhophek) | | -2.59 | -2.41 | 2.25 | 28 | 19.56 | 28 | 56 | 28 |
| G74 | CR Dhan-202 | Aerobic adapted cultivar | 2.96 | -2.61 | 1.92 | 19 | 20.30 | 19 | 38 | 19 |
| G75 | SR-50 | Short rice, landrace from Nagaon, Assam | -0.52 | -0.53 | 5.99 | 113 | 15.00 | 113 | 226 | 113 |
| G76 | MAS 946-1 | Aerobic adapted cultivar | -2.43 | -2.04 | 2.88 | 44 | 17.96 | 44 | 88 | 44 |
| G77 | PB-3 | Pusa basmati | -1.83 | -1.77 | 3.51 | 58 | 17.26 | 58 | 116 | 58 |
| G78 | CR Dhan-201 | Aerobic adapted cultivar | -1.14 | 0.91 | 5.35 | 96 | 13.11 | 96 | 192 | 96 |
| G79 | DRR Dhan-42 | First Drought tolerant MAS derived variety | -2.46 | 2.11 | 2.78 | 42 | 18.14 | 42 | 84 | 42 |
| G80 | DRR Dhan-44 | Aerobic adapted cultivar | -1.47 | 1.31 | 4.55 | 77 | 15.56 | 77 | 154 | 77 |
| G81 | NPS-24 | Wild introgression lines | -1.26 | -1.12 | 4.92 | 88 | 14.50 | 88 | 176 | 88 |
| G82 | NPS-53 | (Swarnax Oryza | -2.47 | -2.08 | 2.75 | 41 | 18.14 | 41 | 82 | 41 |
| G83 | NPS-25 | nivara) | -2.48 | -2.16 | 2.72 | 40 | 18.40 | 40 | 80 | 40 |
| G84 | DB-5 | • | -1.91 | -1.82 | 3.34 | 53 | 17.49 | 53 | 106 | 53 |
| G85 | DB-6 | | -1.08 | 0.81 | 5.57 | 101 | 12.48 | 101 | 202 | 101 |
| G86 | DB-7 | | -0.77 | 0.59 | 5.9 | 110 | 18.00 | 110 | 220 | 110 |
| G87 | DB-9 | | 2.81 | -2.49 | 2.11 | 25 | 19.91 | 25 | 50 | 25 |
| G88 | NPK-13 | | -2.32 | -1.87 | 3.23 | 51 | 17.55 | 51 | 102 | 51 |
| G89 | NPK-27 | | -1.67 | -1.49 | 4.34 | 71 | 16.21 | 71 | 142 | 71 |

Padmashree et al.; Int. J. Environ. Clim. Change, vol. 13, no. 7, pp. 10-23, 2023; Article no.IJECC.99288

| SI. No. | Rice lines | Special Characters | IPCA1 | IPCA2 | ASV | Rank ASV | Mean Yield/plant | Rank of mean | GSI | Rank of GSI |
|---------|--------------|---|-------|-------|-------|----------|---------------------|--------------|-----|----------------|
| G90 | NPK-40 | | -1.88 | -1.79 | 3.41 | 55 | 17.46 | 55 | 110 | 55 |
| G91 | NPK-41 | | 2.31 | -1.86 | 3.26 | 52 | 17.50 | 52 | 104 | 52 |
| G92 | NPK-43 | | -1.17 | -0.94 | 5.3 | 94 | 13.65 | 94 | 188 | 94 |
| G93 | NPK-45 | | 3.15 | 2.84 | 1.13 | 7 | 22.12 | 7 | 14 | 7 |
| G94 | SM-363 | Mutants of N22 | 2.91 | -2.57 | 2 | 21 | 20.24 | 21 | 42 | 21 |
| G95 | SM-669 | | 1.75 | -1.59 | 3.7 | 65 | 16.59 | 65 | 130 | 65 |
| G96 | SM-686 | | -2.58 | 2.39 | 2.33 | 30 | 19.53 | 30 | 60 | 30 |
| G97 | PB-4 | Pusa basmati | 2.86 | -2.54 | 2.05 | 23 | 20.09 | 23 | 46 | 23 |
| G98 | PB-5 | Pusa basmati | 2.61 | -2.42 | 2.16 | 27 | 19.67 | 27 | 54 | 27 |
| G99 | Sahbhagidhan | Drought tolerant variety | -1.48 | 1.33 | 4.5 | 76 | 15.68 | 76 | 152 | 76 |
| G100 | Sabita | O <i>ryza sativa</i> cultivar | -0.79 | -0.62 | 5.85 | 108 | 10.04 | 108 | 216 | 108 |
| G101 | DRR Dhan-41 | Drought tolerant variety | -1.07 | -0.79 | 5.6 | 102 | 11.48 | 102 | 204 | 102 |
| G102 | KMR-3 | Variety | -0.86 | -0.7 | 5.8 | 105 | 11.12 | 105 | 210 | 105 |
| G103 | IR-64 | Cultivated Mega variety | 1.28 | -1.14 | 4.86 | 86 | 14.73 | 86 | 172 | 86 |
| G104 | ATR-486 | Introgression line (Azucena × Dular) | -1.18 | 0.96 | 5.1 | 93 | 13.70 | 93 | 186 | 93 |
| G105 | ASG-73 | Landrace from West Bengal | 3.11 | 2.68 | 1.86 | 17 | 20.46 | 17 | 34 | 17 |
| G106 | ASG-126 | Landrace collected from Uttar Pradesh | -1.73 | 1.42 | 3.73 | 66 | 16.45 | 66 | 132 | 66 |
| G107 | Saali | Oryza glaberrima | 1.68 | -1.53 | 4.28 | 70 | 16.25 | 70 | 140 | 70 |
| G108 | Dissi | accessions | 1.24 | -1.09 | 4.95 | 89 | 14.40 | 89 | 178 | 89 |
| G109 | Mow | | -2.34 | -1.9 | 3.23 | 50 | 17.59 | 50 | 100 | 50 |
| G110 | Mouli | | 2.52 | -2.2 | 2.56 | 37 | 18.90 | 37 | 74 | 37 |
| G111 | Basmathi-370 | Traditional Bamati cultivar | -2.58 | 2.37 | 2.29 | 29 | 19.54 | 29 | 58 | 29 |
| G112 | Thurur Bhog | Landrace | -0.31 | 0.44 | 6.354 | 117 | 11.00 | 117 | 234 | 117 |
| G113 | D-92 | North-Eastern Landrace | 2.98 | -2.62 | 1.89 | 18 | 20.42 | 18 | 36 | 18 |
| G114 | JBB-661 | Tropical japonica × | -2.84 | 2.51 | 2.08 | 24 | 19.92 | 24 | 48 | 24 |
| G115 | JBB-610 | indica introgressed | 1.78 | -1.71 | 3.6 | 62 | 16.91 | 62 | 124 | 62 |
| G116 | JBB-684 | lines | 2.42 | -2.01 | 2.95 | 46 | 17.90 | 46 | 92 | 46 |
| G117 | JBB-1325 | | 3.21 | -0.98 | 0.89 | 3 | 23.90 | 3 | 6 | 3 |
| G118 | JBB-631-1 | | -3.21 | -2.14 | 0.75 | 2 | 24.70 | 2 | 4 | 2 |

ASV-AMMI stability value, GSI- Genotypic selection index, IPCA- interaction principal component axes

Table 2. AMMI ANOVA for Grain yield

| Source | d.f. | Sum of squares | Mean squares | F ratio | %TSS | G*E% |
|-----------|--------|----------------|--------------|---------|-------|-------|
| ENV | 3.00 | 79528.20 | 26509.40** | 3.18 | 1.96 | |
| GEN | 117.00 | 1010908.98 | 8640.24** | 1.03 | 25.02 | |
| ENV*GEN | 351.00 | 2948569.95 | 8400.48** | 1.00 | 73.00 | |
| PC1 | 119.00 | 2942036.09 | 24722.99** | 2.96 | 99.77 | 79.20 |
| PC2 | 117.00 | 6128.92 | 52.38 | 0.006 | 0.20 | 15.18 |
| Residuals | 944.00 | 7855141.49 | 8321.12 | NA | 0 | |
| Mean | 17.100 | | | | | |
| SD | 3.327 | | | | | |
| Variance | 11.070 | | | | | |
| CV (%) | 19.457 | | | | | |
| CD (%) | 12.527 | | | | | |

^{**}Significance @ p=0.001, %TSS= % of total sum of squares, IPCA= interaction principle component Axes, %G*E= % of genotype and environment interactions, SD= standard deviation, CV=coefficient of variation, CD= critical difference

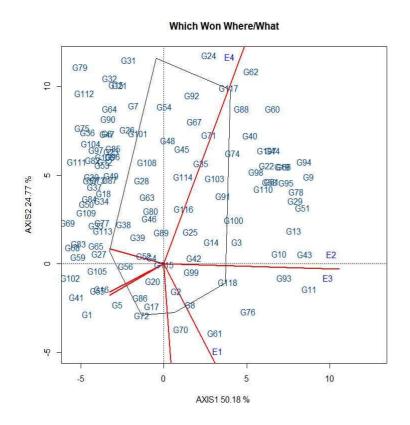
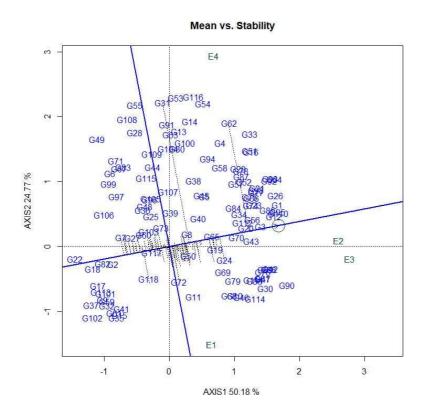
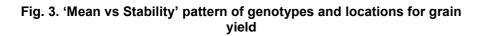


Fig. 1. 'Which won-where' pattern of genotypes and locations for grain yield

Discrimitiveness vs. representativenss E4 G55 2 G62 G33 G44 AXIS2 24.77 % E2 0 G24 E3 E1 0 2 3 -1 AXIS1 50.18 %

Fig. 2. 'Discrimitiveness vs representativeness' pattern of genotypes and locations for grain yield





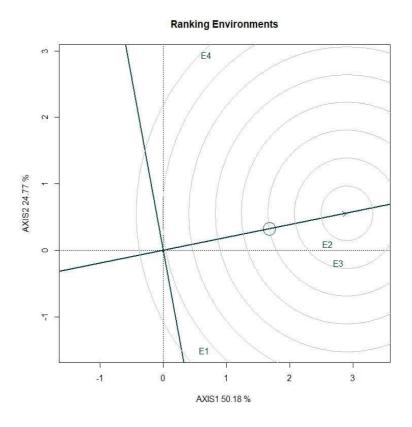


Fig. 4. 'Ranking of Environments' for grain yield

studied trait. In our investigation, the mean vs. stability analysis recorded Phouren, Chakhao, Moirangphou-Yenthik, and Langphou-Chakao a high mean yield performance, whereas Phouren, JBB-1325 and Swarna were found to be the most stable lines

The GGE biplot for grain yield per hectare for ranking environments (Fig. 4) indicated that the environment ICAR-IIRR-Dry 2019 (E2) and ICAR-IIRR-Wet 2020 (E3) formed smaller angle and contemplated representative environments for grain yield per plant than other environments, therefore these environments are more informative. In terms of line discrimination, ICAR-IIRR-Dry 2020 (E4) had the longest vector and formed the smallest angle with AEA, followed by ICAR-IIRR-Dry 2019 (E2) and ICAR-IIRR-Wet 2020 (E3), Dhadesugur -Wet 2020 (E1) has the shortest vector forms the biggest angle with AEA, and thus environment is not suitable for line discrimination but can be used after culling unstable lines. The present findings of significant sources of variation have been previously noted in rice [22,23] and other cereal crops [24]. Similarly, to our results Phouren, NPK-43, JBB-684 and Ratnamudi were identified as high yielding lines under conditions [14]. Although in the present study, both the AMMI model and GGE biplots have been found to be useful tools for understanding GE interactions, however, GGE biplots may provide more meaningful information with regard to genotype-environment relationships.

4. CONCLUSION AND RECOMMENDA-TIONS

A significant Genotype × Environment interaction was detected for yield by evaluating 118 lines across four environments using AMMI analysis. The combined analysis of variance (ANOVA) manifested significant variations for the tested rice lines, locations, years, genotype x year, and genotype x location interactions revealing the influence of environmental factors on yield per plant. ΑII Four environments showed discrimination power, whereas E2 and E3 were found as the representative environment. The AMMI biplot showed that PC1 contributed 79.20% variability and PC2 contributed 15.18% variability (Table 2). From the GGE biplot analysis, the rice lines Phouren, JBB-631-1 and JBB-1325, PUP 229, TI-112, Longphou, NPK-45, Moirangphou-Yenthink, TI-3, and Phouren-Amubi have top-ranked rice lines based on ASV-AMMI stability value. The rice lines Phouren, PUP-229,

and TI-112 own the first sub-group which is Dhadesugur-Wet 2020 (E1). The rice lines Langphou, NPK-45 won the second sub-group which was produced by ICAR-IIRR-Wet 2020 (E3). Environment ICAR-IIRR-Dry 2019 (E2) was the third subgroup the rice lines Moirangphou-Yenthik and TI-3 were the winner. The ICAR IIRR-Dry 2020 (E4) environment formed the fourth subgroup where Phouren-Amubi, TI-128 and JBB-1325 were the winners. The high-yielding stable lines can be recommended for release after evaluation at multiple locations and can also be deployed in breeding programs for enhancing yield per plant under irrigated ecosystems.

ACKNOWLEDGEMENTS

The authors acknowledge Drs. C N Neeraja (Principal Scientist), Suneetha Kota (Senior Scientist), Satendra Kumar Mangrauthia (Senior Scientist) from ICAR-IIRR for sharing the seeds of experimental plant materials. The authors acknowledge Drs Brajendra and Vakada Manasa for soil field analysis. The authors are grateful to Director, **ICAR-IIRR** for the providing laboratory and field research facilities. The authors are thankful to the Head, ARS Dhadesugur, and UAS, Raichur for providing field facilities.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Islam MR, Sarker MR, Sharma N, Rahman MA, Collard BC, Gregorio GB, Ismail AM. Assessment of adaptability of recently released salt tolerant rice varieties in coastal regions of South Bangladesh. Field Crops Res. 2016;1:190:34-43.
- Balakrishnan D, Subrahmanyam D, Badri J, Raju AK, Rao YV, Beerelli K, Mesapogu S, Surapaneni M, Ponnuswamy R, Padmavathi G, Babu VR. Genotypex environment interactions of yield traits in backcross introgression lines derived from Oryza sativa cv. Swarna/Oryza nivara. Front. plant sci. 2016;19:7:1530.
- 3. Finlay KW & Wilkinson GN. The analysis of adaptation in a plant-breeding programme. Aust. J. Agric. Res. 1963; 14(6):742-54.

- 4. Eberhart ST & Russell WA. Stability parameters for comparing varieties 1. Crop sci. 1966;6(1):36-40.
- 5. Perkins J & Jinks. Environmental and genotype-environmental components of variability. VI. Diallel sets of crosses. Heredity. 1968;25:29-40.
- 6. Freeman GH & Perkins JM. Environmental and genotype-environmental components of variability VIII. Relations between genotypes grown in different environments and measures of these environments. Heredity. 1971;27(1):15-23.
- 7. Shukla GK. Some statistical aspects of partitioning genotype environmental components of variability. Heredity. 1972; 29(2):237-45.
- 8. Kang MS. Simultaneous selection for yield and stability and yield-statistic. J. Agron. 1993;85(3):754-7.
- 9. Kempton RA. Adjustment for competition between varieties in plant breeding trials. The J. Agril. Sci. 1984;98(3):599-611.
- Crossa J, Cornelius PL. Sites regression and shifted multiplicative model clustering of cultivar trial sites under heterogeneity of error variances. Crop Sci. 1997;37(2): 406-15.
- 11. Gauch Jr HG & Zobel RW. Identifying mega-environments and targeting genotypes. Crop sci. 1997;37(2):311-26.
- 12. Gauch Jr HG. Model selection and validation for yield trials with interaction. Biometrics. 1988;1:705-15.
- Yan W, Hunt LA, Sheng Q, Szlavnics Z. Cultivar evaluation and mega-environment investigation based on the GGE biplot. Crop Sci. 2000;40(3):597-605.
- Padmashree R, Barbadikar KM, Magar 14. ND, Balakrishnan D, Channappa G, Siddaiah Madhav MS. AM. Bharamappanavara M, Phule AS, Diwan J. Sundaram RM. Genome-wide association studies in rice germplasm reveal significant genomic regions for root and yield-related and traits under aerobic irrigated conditions. Frontiers in Plant Science. 2023;14:1588.
- Purchase JL, Hatting H, Van Deventer CS. Genotypex environment interaction of winter wheat (*Triticum aestivum* L.) in South Africa: II. Stability analysis of yield performance. S. Afr. J. Plant Soil. 2000;1:17(3):101-7.

- 16. Babu PM, Neeraja CN, Rathod S, Suman K, Uttam GA, Chakravartty N, Lachagari VR, Chaitanya U, Rao LV, Voleti SR. Stable SNP allele associations with high grain zinc content in polished rice (Oryza sativa L.) identified based on ddRAD sequencing. Front. Genet. 2020;11: 11:763.
- Sharifi P, Aminpanah H, Erfani R, Mohaddesi A, Abbasian A. Evaluation of genotypex environment interaction in rice based on AMMI model in Iran. Rice sci. 2017 May 1;24(3):173-80.
- 18. Ashwini GL, Rajanna MP, Deepak CA, Chethana BS, Shobha D, Gouda TH, Dushyanthkumar BM, Hanamaratti NG, Ramesh S, Nagaraj MS, Mahantashivayogayya KH. Stability analysis for grain yield and quality traits in selected traditional and improved varieties of rice over different Zones of Karnataka. Oryza. 2019;56(2):193-203.
- Dewi AK, Chozin MA, Triwidodo H, Aswidinnoor H. Genotypex environment interaction, and stability analysis in lowland rice promising genotypes. Int. J. Agron. Agril. Res. 2014;5(5):74-84.
- 20. Oladosu Y, Rafii MY, Abdullah N, Magaji U, Miah G, Hussin G, Ramli A. Genotypex Environment interaction and stability analyses of yield and yield components of established and mutant rice genotypes tested in multiple locations in Malaysia. Acta Agriculturae Scandinavica, Section B—Soil & Plant Sci. 2017;3:67(7):590-606.
- Khan MM, Rafii MY, Ramlee SI, Jusoh M, 21. Al Mamun M. AMMI and GGE biplot analysis for yield performance and stability assessment of selected Bambara groundnut (Vigna subterranea L. Verdc.) genotypes under the multi-environmental Sci. trials (METs). Rep. 2021:23: 11(1):22791.
- 22. Mwiinga B, Sibiya J, Kondwakwenda A, Musvosvi C, Chigeza G. Genotype x environment interaction analysis of soybean (*Glycine max* (L.) Merrill) grain yield across production environments in Southern Africa. Field crops Res. 2020;1: 256:107922.
- 23. Shrestha S, Asch F, Dusserre J, Ramanantsoanirina A, Brueck H. Climate effects on yield components as affected by genotypic responses to variable environmental conditions in upland rice

systems at different altitudes. Field Crops Res. 2020;12: 134:216-28.

24. Shojaei SH, Mostafavi K, Omrani A, Omrani S, Nasir Mousavi SM, Illes A,

Bojtor C, Nagy J. Yield stability analysis of maize (*Zea mays* L.) hybrids using parametric and AMMI methods. Scientifica. 2021;25:1-9.

© 2023 Padmashree et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
https://www.sdiarticle5.com/review-history/99288