



# Molecular Detection of Some Virulence Genes and Determination the Inhibitory Efficiency of Nano Hybrid Antibiotic against (*Escherichia coli*) Isolated from Urinary Tract Infection

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## Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

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## ABSTRACT

**Objective:** *Escherichia coli* is considered the most common microorganism in pathological samples related to urinary tract infections especially in women and children. It contains many virulence factors helping in pathogenicity However it is sensitive to fosfomycin that has broad spectrum in its activity against bacteria which its activity also could be confirmed using nanobiotechnology.

**Methods:** The study included 25 isolates of *E. coli* from patients with urinary tract infections. DNA extracted and PCR technique used for detection of 3 genes (*afa*, *pap* and *fimh*) coding for virulence factor afimbrial adhesions proteins, pilus rod and Type 1 fimbriae respectively. Magnesium oxide was used as a carrier in intercalating of fosfomycin between its layers. fourier transform infrared

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spectroscopy (FT-IR) atomic force microscope (AFM) and X-ray diffraction (XRD) were used for characterizing of prepared nanohybrid antibiotic. Antibacterial activity of against *E. coli* has been tested.

**Results:** PCR results showed bands with 750, 328 and 508 bps for the three genes *afa*, *pap* and *fimh* respectively. The all of three techniques used to confirm the hydrating process indicated formation of new compound through the appearance of new group of spectrums in FT-IR, formation of spherical nanoparticles in AFM and appearance of a new diffraction planes in XRD.

**Conclusion:** It can be concluded that *E. coli* isolate under study were virulent due the presence of virulence genes. Using nanotechnology, Fosfomycin can be used to prepare nanohybrid antibiotic by loading on Magnesium oxide. The prepared nano-particle was efficient in inhibiting *E. coli* isolated from urinary tract infections.

**Keywords:** Characterization; *Escherichia coli*; fosfomycin; nanohybrid; nanohybrid antibiotic; urinary tract infections.

## 1. INTRODUCTION

Urinary tract infections are one of the most common types of bacterial infections, with an estimated 400 million cases and 230,000 deaths globally in 2019. 50% of women suffer from this type of infection at least once in their lives, and of course recurrence of the infection is common. The rate of infection increases in the elderly, as in hospitals the highest rate of urinary tract infections is recorded among patients over the age of 65, after urinary system infections [1,2].

Gram-negative bacteria account for the majority of urinary tract infections, although gram-positive pathogens are also responsible. Uncomplicated UTIs are monobacterial in >95% of cases. The frequent cause of uncomplicated UTIs is *E. coli*, followed by, *Klebsiella pneumoniae*, *Staphylococcus saprophyticus*, *Enterococcus faecalis*, *Proteus mirabilis*, and group B streptococci [3]. The uropathogenic *Escherichia coli* (UPEC) is considered as the main cause of this type of infection, accounting for more than 75% of uncomplicated and 65% of complex infections in the urinary tract [2]. Several virulence factors are involved in its uropathogenesis, including those that encode proteins of the fimbrial surface adhesins family, such as *Fimh*, *Afa* and *Pap* [4]. *The Afa (afimbrial adhesin) and Pap (pyelonephritis-associated pili) operons are the most common adhesion mediators. They facilitate adhesion of E. coli to uroepithelial cells and protect them from being washed away by urine flow increasing their pathogenesis* [5]. *fimh* (fim stand for of fimbriae), by its specificity for D-mannose-containing structures on host cells, facilitate binding of the fimbriae to host receptors [2].

Fosfomycin is one of the most important broad-spectrum bactericidal antibiotics used for

treatment of UTI and multidrug-resistant bacterial infection [6,7].

Nanotechnology has recently brought about a global technical revolution through the manufacture of new nanomaterials with advanced physical, chemical and biological properties. Nanoantibiotics are one of those materials that have captured the attention of researchers due to their reduced toxicity and the possibility of using them to inhibit drug-resistant microorganisms, in addition to their low cost compared to traditional antibiotics [8].

Magnesium oxide is one of the important metal oxides due to its unique and excellent properties, including optical, electrical, thermal, mechanical, and chemical properties, as well as its ionic properties and its antimicrobial activity as radical oxygen species (ROS) generation. At the nanoscale level, magnesium oxide shows high effectiveness due to it containing a large number of highly effective edges, the nature of its surface structure and its unnatural cross-linked levels, as well as its surface area-to-volume capacity [9,10].

The current study aimed to prepare an efficient nanohybrid antibiotic using fosfomycin inhibiting virulence UTI isolate.

## 2. MATERIALS AND METHODS

### 2.1 DNA Extraction and Gene Detection

Twenty-five uropathogenic *E. coli* isolates were obtained from department of biology – college of science- university of Kerbala. Addbio bacterial DNA extraction kits were used for extraction of DNA from the isolates according to the manufacturer instruction

([http://addbio.net/default/subc/c03.php?com\\_board\\_basic=read\\_form&com\\_board\\_idx=47&&com\\_board\\_search\\_code=&com\\_board\\_search\\_value1=&com\\_board\\_search\\_value2=&com\\_board\\_page=&&com\\_board\\_id=25&&com\\_board\\_id=25](http://addbio.net/default/subc/c03.php?com_board_basic=read_form&com_board_idx=47&&com_board_search_code=&com_board_search_value1=&com_board_search_value2=&com_board_page=&&com_board_id=25&&com_board_id=25)). Primers of gene under study were taken from Mishra and Panda [5] study. DNA purity was checked by Nano-drop. Virulence genes under study are shown in Table 1.

The amplification conditions for the virulence genes included one cycle for initial denaturation by 95°C for 5 min. Denaturation step included 35 sec in 95°. Annealing temperatures for *afa*, *pap* and *fimh* were 55, 57 and 60°C respectively and for one min for 35 cycle, and extension and final extension steps included one cycle in 72°C for 60 sec and 7 min respectively.

## 2.2 Preparation of Nanohybrid Fosfomycin

The nanohybrid Fosfomycin was prepared according to the method that described by Bashi et al. [11] as follows:

- a- **Preparation of Fosfomycin Solution:** This solution was prepared by dissolving 1.2 gm of Fosfomycin in 40 ml of suitable solvent and the volume completed to 50 ml with same solvent, too.
- b- **Preparation of Carrier:** Carrier solution was prepared by dissolving 1 gm of magnesium oxide (MgO) in 50 ml of 50% ethanol.

“Briefly, fifty ml of Fosfomycin solution were added drop by drop to the MgO solution with stirring. The mixture was stirred by magnetic stirrer at room temperature for 24 hrs. and the mixture was placed in an incubator at 40°C for 18 hrs. The precipitate was separated by

centrifuge at 5000 rpm for 20 min, washed with deionized water for several times and was dried at 50°C. Finally, the dried precipitate was grinded well and gave the symbol Fos-MgO” [12].

## 2.3 Characterization of Synthesized Nanohybrid Fosfomycin

The nanohybrid Fosfomycin of present study was characterized by using several methods according to [13,14] as follows:

- (i) **Fourier Transform Infrared Spectroscopy (FT-IR):** The infrared spectrum study for each of nanohybrid Fosfomycin and Fosfomycin free form as well as MgO was carried out by making disks through mixing with potassium bromide (KBr) after grinding well. The infrared spectrum was measured in a wave number range (4000-400) cm<sup>-1</sup>.
- (ii) **X-ray Diffraction Spectrum (XRD):** The nanohybrid Fosfomycin of the present study was characterized using XRD which explains the difference in the layer before and after the intercalating process by using Bragg’s law.
- (iii) **Atomic Force Microscope (AFM):** The current study included measuring the diameter, size and aggregation of the nanohybrid Fosfomycin nanoparticles by using AFM.

## 2.4 Detection of Inhibitory Activity of Free and Nanohybrid Fosfomycin

The inhibitory activity of free and nanohybrid form of Fosfomycin was detected against *Escherichia coli* isolated from urinary tract infections according to the well diffusion method [15] by using antibiotic concentration of (16-512) µg/ml.

**Table 1. Virulence genes primers and product size [5]**

No.	Virulence genes	Oligonucleotide sequence (5'-3') forward and reverse	Size of amplicons
1.	<i>afa</i>	F-GCTGGGCAGCAAAGTATAACTCTC R-CATCAAGCTGTTTGTTCGTCGCCG	750 bp
2.	<i>Pap</i>	F-GACGGCTGTAAGTGCAGGGTGTGGCG R- ATATCCTTTCTGCAGGGATGCAATA	328 bp
3.	<i>Fimh</i>	F-TGCAGAACGGATAAGCCGTG R-GCAGTCACCTGCCCTCCGGTA	508 bp

### 3. RESULTS AND DISCUSSION

#### 3.1 Molecular Detection of *afa*, *pap* and *fimh* Genes

The results in following images show the appearance of *afa* bands with a molecular size of 750 bp, *pap* bands with a molecular size of 328 bp and *fimh* bands with a molecular size of 508

bp, in 6 and 10 and 6 of samples respectively. Table 2 show appearance of virulence genes in specimens.

The genes *afa*, *pap* and *fimh* were presented in percentage of (24, 40 and 24%) respectively which indicate *pap* is the most predominant gene in the urinary tract infection isolates under study (Table 2).



Fig. 1. Gel electrophoresis result of *afa* gene (750 bps), were (M) indicate DNA Ladder 1500 bps



Fig. 2. Gel electrophoresis result of *pap* gene (328 bps), were (M) indicate DNA Ladder 1500 bps

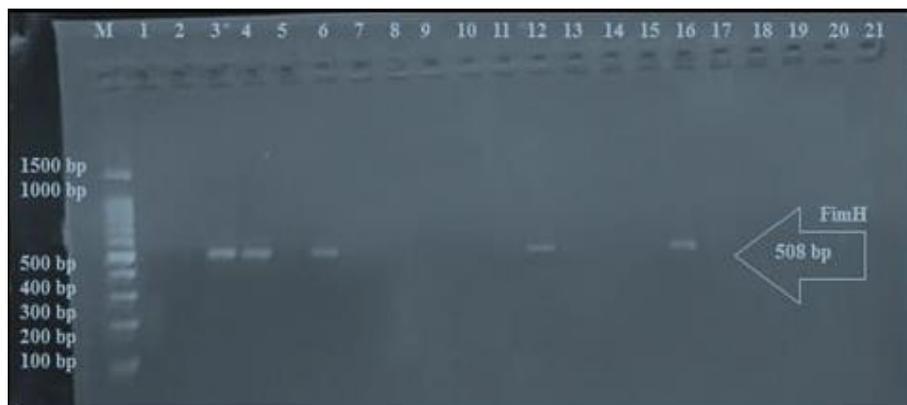


Fig. 3. Gel electrophoresis result of *fimh* gene (508 bps), were (M) indicate DNA Ladder 1500 bps

**Table 2. Appearance of virulence genes in specimens**

Sp. No. Gene	S1	S2	S3	S4	S5	S6	S7	S8	S9	S10	S11	S12	S13	S14	S15	S16	S17	S18	S19	S20	S21	S22	S23	S24	S25	
<i>afa</i>	.	+	.	.	+	.	+	.	.	+	+	.	+	.	.	.	.	.	.	.	.	.	.	.	.	.
<i>pap</i>	.	+	+	.	+	.	+	.	+	.	+	.	.	+	.	.	.	.	.	.	.	.	.	+	+	.
<i>fimh</i>	.	.	+	+	.	+	.	.	.	.	.	+	.	.	.	+	.	.	.	.	.	.	.	+	.	.

Many virulence factors have a role in the pathogenicity of *Escherichia coli* in urinary tract infection of which adhesion proteins have a prime function. Type 1 fimbriae are encoding result of the *fim* operon cluster and consist of a prime protein, FimA, associated with supplementary proteins *fimG* and *fimF* and the adhesion protein *fimH*. *pap* gene cluster is an aggregation of 11 genes that encode for main element of the pilus rod. The five defined *afaA* to *E* genes encode as known afimbrial adhesions proteins have an important role in the pyelonephritis. *afa* operon encode for proteins act as specific binding actor to human erythrocyte receptors and uroepithelial cells [16].

### 3.2 Characterization of Nanohybrid Antibiotics

#### 3.2.1 Characterization by FT-IR

Fig. 4 shows the results of FT-IR spectrum of free Fosfomycin. The broad band that appeared at (3182)  $\text{cm}^{-1}$  was attributed to a stretch vibration of the two hydroxyl groups. The appearance of the band at frequency of (2939)  $\text{cm}^{-1}$  was due to the aliphatic (C-H) stretch. The

band which appeared at 1633  $\text{cm}^{-1}$  includes the stretch of P=O group. In addition, the appearance of the two bands at (1139 and 1076)  $\text{cm}^{-1}$  indicate the stretch of etheric C - O - C in the epoxide ring [13].

Fig. 5 shows few bands of absorption in the FT-IR spectrum of magnesium oxide (MgO). The band at 3437  $\text{cm}^{-1}$  is related to O-H group's stretching vibration which associated to the physically adsorbed water, while the band at 1514  $\text{cm}^{-1}$  related to the bending vibration of O-H group surface of the mentioned absorbed water. Anyhow the band at 1429  $\text{cm}^{-1}$  is related to the asymmetric stretching of ( $\text{CO}_3^{2-}$ ) and the vibration of metal bond was showed at (680-580)  $\text{cm}^{-1}$  [17].

Fig. 6 shows a combination FT-IR spectrum of both the Fosfomycin and MgO carrier. The broad band at 3414  $\text{cm}^{-1}$  which undergoes shift to high frequency is due to the stretch of two hydroxyl groups. The results showed a shift to P=O stretch group to high frequency as it appeared at 1649  $\text{cm}^{-1}$ . Finally, the two bands that belong to stretch of C-O-C in the epoxide ring underwent a shift to low frequencies as they appeared at (1107 and 1003)  $\text{cm}^{-1}$  [13].

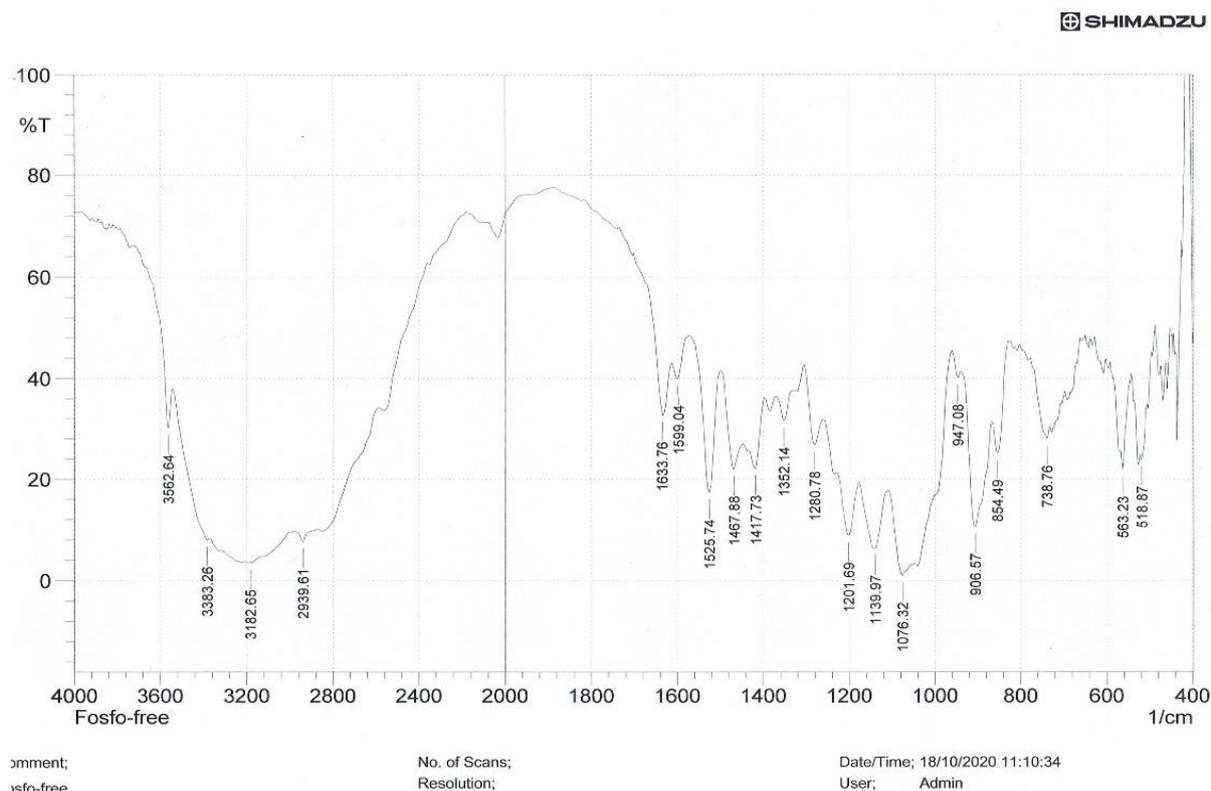


Fig. 4. FT-IR spectrum of free Fosfomycin

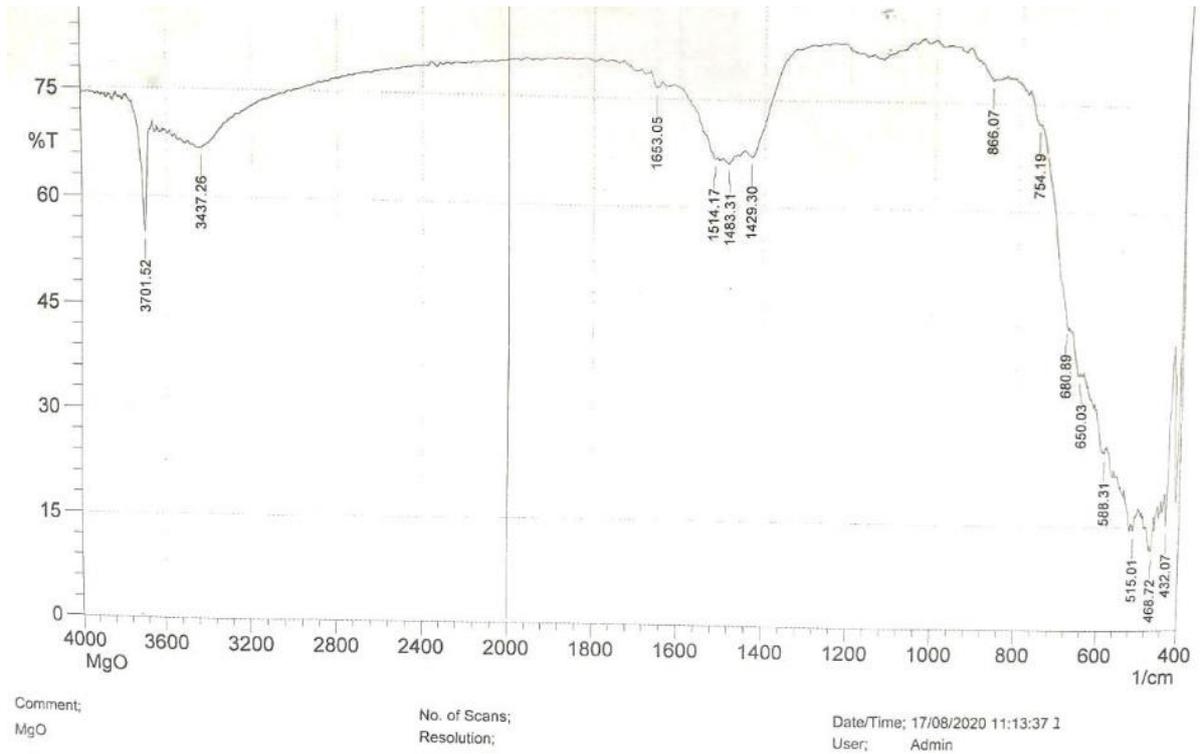


Fig. 5. FT-IR spectrum of Magnesium oxide

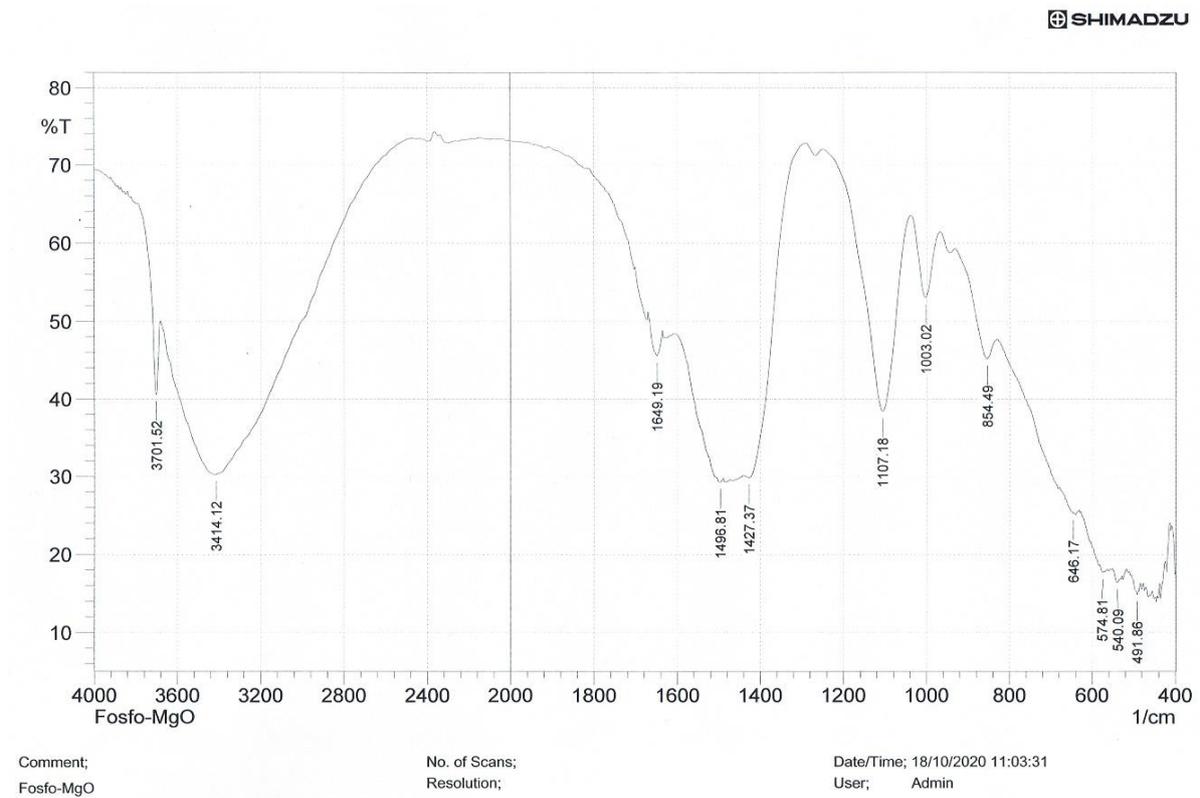


Fig. 6. FT-IR spectrum of nanohybrid Fosfomycin (Fos-MgO)

### 3.2.2 Characterization by XRD

In current study diffraction spectrum of X-ray of MgO alone as well as Fos-MgO were measured to obtain the differences in the layer thickness before and after loading of Fosfomycin on MgO layers by using Bragg's law.

Fig. 7 shows XRD analysis of MgO. As shown, many diffraction planes are appeared in the spectrum, as follows: At 37.95°, (111) with crystalline distance of 0.23 nm, at 42.95°, (200) with crystalline distance of 0.21 nm, at 62.37°, (220) with crystalline distance of 0.14 nm, at

74.75°, (311) with crystalline distance of 0.126 nm and at 78.66° the diffraction plane was (222) with crystalline distance of 0.121 nm [17].

Fig. 8 shows the XRD analysis after the intercalation between Fosfomycin and MgO layers as follows: crystalline distance of 0.49 nm was for diffraction plane (003) which appeared at 17.95° while crystalline distance of 0.23 nm showed for another plane (006) at 37.56°. The XRD results provided a convenient indication for the intercalation between Fosfomycin and magnesium oxide to compose Fos-MgO.

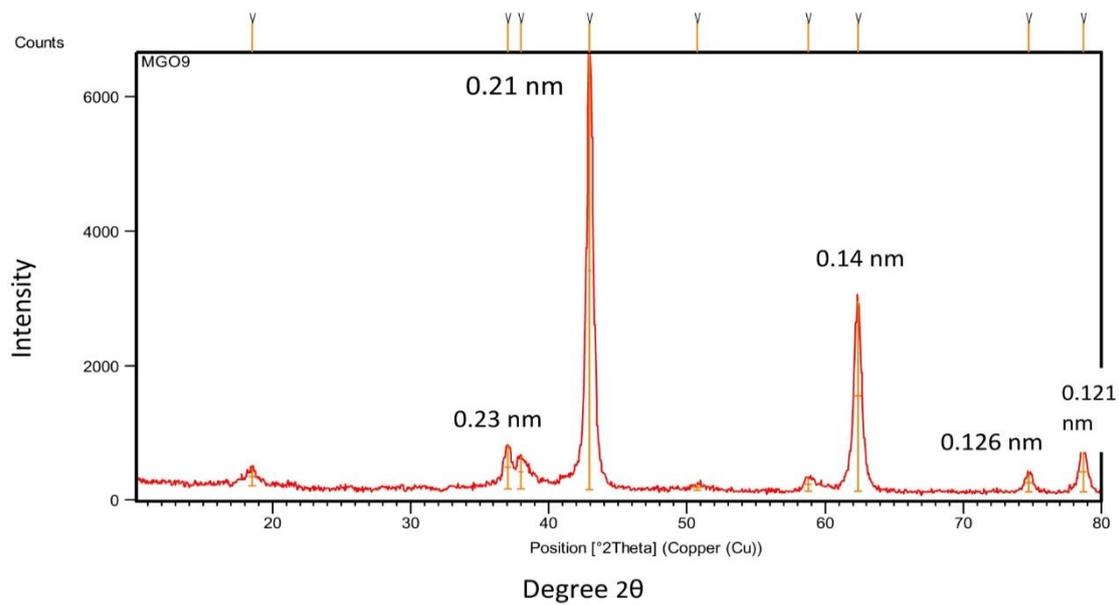


Fig. 7. XRD spectrum of Magnesium oxide

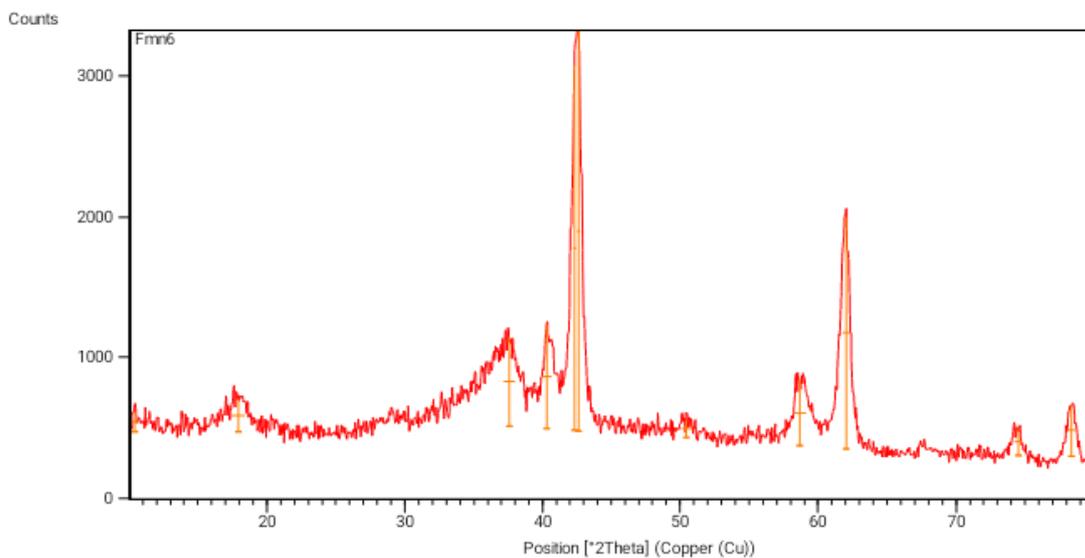


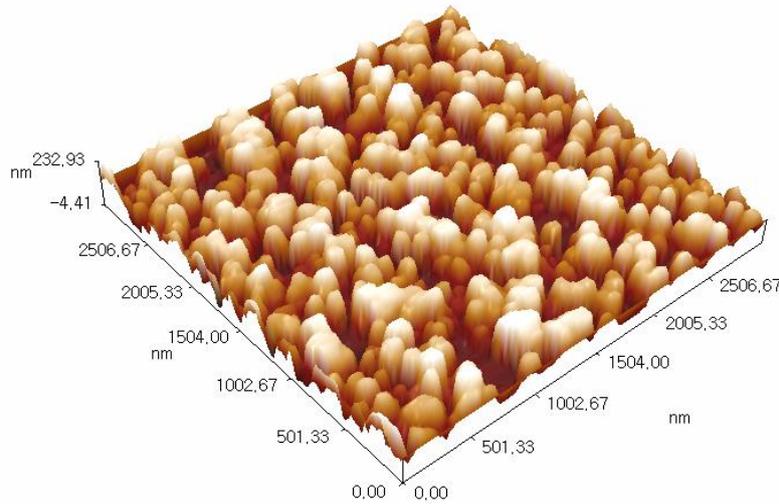
Fig. 8. XRD spectrum of nanohybrid Fosfomycin (Fos-MgO)

### 3.2.3 Characterization by AFM

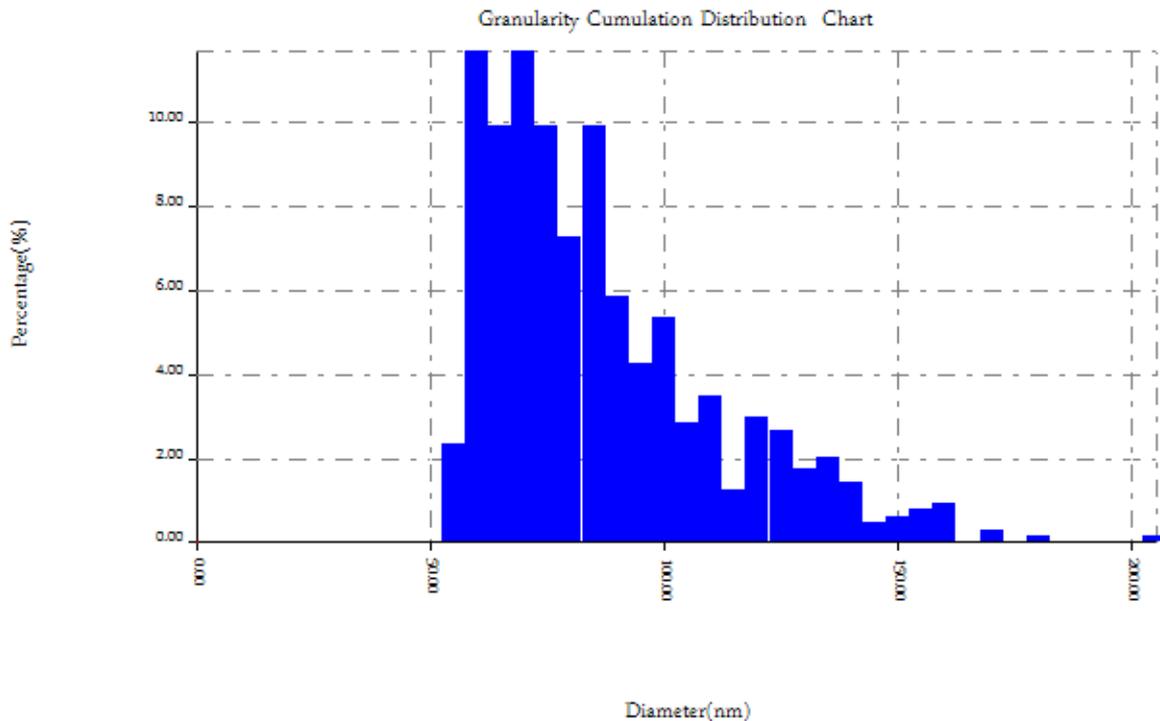
The outer surface of Fos-MgO was scanned by atomic force microscope (AFM).

Fig. 9 show the three-dimensional section of prepared nano antibiotic. The nanoparticles were almost spherical in shape with height of 232 nm. Fig. 10 showed the chart of granularity

cumulation distribution of Fos-MgO hybrid. Nanoparticles with highest percentage (11.65%) was belonged to diameters of (60 and 70) nm. Fos-MgO hybrid showed particle size means about of 84.3 nm. These results were different from those obtained by Jabur [10] who obtained a diameter average size of 77 nm for the prepared nanohybrid ciprofloxacin loaded on MgO.



**Fig. 9. Three-Dimensional image of nanohybrid Fosfomycin (Fos-MgO) by Atomic Force Microscope**



**Fig. 10. Granulity Cumulation Distribution Chart of nanohybrid Fosfomycin (Fos-MgO) by Atomic Force Microscope**

### 3.2.4 The inhibition activity of the nanohybrid antibiotic against *E. coli*

Fosfomycin is an antibiotic with a broad-spectrum activity and inhibit cell wall synthesis by interfering with the formation of UDP N-acetylmuramic acid (the peptidoglycan precursor) [18,19]. The inhibition activity of free and the nanohybrid antibiotic (Fos-MgO) was studied against ten isolates of *E. coli* isolated urinary tract infections. Table 3 shows the inhibition activity of free Fosfomycin. The diameters size indicates increasing of inhibition by increasing of concentrations. Using 512 µg/ml of Fosfomycin the diameter size of inhibition varied from 22 to 26.6 mm for *E. coli* 6 and *E. coli* 1 respectively. On the other hand, Table 3 show that Fos-MgO nanohybrid with concentration of 512 µg/ml gave large inhibition diameter of 26.5 mm on *E. coli* 1 isolate and least inhibition diameter of 22 mm on *E. coli* 6 isolate.

By comparing the diameter of the inhibition zone for both the free and hybrid antibiotics with

magnesium oxide on the same isolate in Tables 3 and 4, it is noted that the diameter of inhibition for the free antibiotic is larger than the nano-hybrid antibiotic for the same concentration, but it must also be noted that in the case of nano-hybrid, the percentage of antibiotic loaded on the carrier is approximately one-third of the percentage of free antibiotic used for inhibition.

Many studies applied to comparing between the inhibitory activity of free antibiotics and that of nanohybrid one against pathogenic bacteria. In study conducted by Al-Fatlawi [20] it has been shown that the nanohybrid antibiotic was very efficient in inhibiting of isolates. However, she studied the effect of azithromycin of *Klebsiella pneumonia* isolates. Also, our results were similar to those announced by Kumar et al. [21] where found the high efficiency of doxycycline nanoparticle in the inhibiting of *E. coli* isolates compared to that of free doxycycline.

**Table 3. Inhibition activity of free Fosfomycin**

No.	Antibiotic concentration Mg/ml	16	32	64	128	256	512
	<b>Bacterial isolate</b>						
1.	<i>Escherichia coli</i> 1	0	11	13	17.5	23.5	26.5
2.	<i>Escherichia coli</i> 2	9.5	12	14.5	18	21.5	24
3.	<i>Escherichia coli</i> 3	7.5	9.5	12.5	14.5	16.5	23.5
4.	<i>Escherichia coli</i> 4	0	9	12.5	14	19.5	23
5.	<i>Escherichia coli</i> 5	6	10.5	14	16	19	23.5
6.	<i>Escherichia coli</i> 6	7.5	11	14.5	16	18.5	22
7.	<i>Escherichia coli</i> 7	10	13.5	15	16.5	21	25
8.	<i>Escherichia coli</i> 8	11	13	17	19.5	23	25.5
9.	<i>Escherichia coli</i> 9	8.9	12	16.5	18.8	20.5	23.5
10.	<i>Escherichia coli</i> 10	9	12	17	19	22	24.5

**Table 4. Inhibition activity of nanohybrid Fosfomycin (Fos-MgO)**

No.	Antibiotic concentration Mg/ml	16	32	64	128	256	512
	<b>Bacterial isolate</b>						
11.	<i>Escherichia coli</i> 1	0	0	9	10.5	12.5	15.5
12.	<i>Escherichia coli</i> 2	0	7.5	10.5	12.5	14	16
13.	<i>Escherichia coli</i> 3	0	0	11	13	14	17.5
14.	<i>Escherichia coli</i> 4	0	7	10	11	12.5	14
15.	<i>Escherichia coli</i> 5	0	0	12	13	14.5	15
16.	<i>Escherichia coli</i> 6	0	8	12	14	15.5	18.5
17.	<i>Escherichia coli</i> 7	0	9.5	13	15	16.5	19.5
18.	<i>Escherichia coli</i> 8	0	10	12.5	14	15.5	17.5
19.	<i>Escherichia coli</i> 9	0	0	12	14.5	16.5	19
20.	<i>Escherichia coli</i> 10	0	0	9	10.5	12.5	15

#### 4. CONCLUSION

Nanotechnology has recently brought about a global technical revolution through the manufacture of new nanomaterials with advanced physical, chemical and biological properties. Using nanotechnology, Fosfomycin can be used to prepare nano-hybrid antibiotic by loading on Magnesium oxide. The prepared nano-particle was efficient in inhibiting *E. coli* isolated from urinary tract infections.

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#### COMPETING INTERESTS

Author has declared that no competing interests exist.

#### REFERENCES

1. Russell SK, Harrison JK, Olson BS, Lee HJ, O'Brien VP, Xing X, et al. Uropathogenic *Escherichia coli* infection-induced epithelial trained immunity impacts urinary tract disease outcome. *Nature Microbiology*. 2023;8(5):875-888.
2. Whelan S, Lucey B, Finn K. Uropathogenic *Escherichia coli* (UPEC)-Associated Urinary Tract Infections: The Molecular Basis for Challenges to Effective Treatment. *Microorganisms*. 2023;11(9):2169.
3. Sobel JD, Kaye D. *Principles and Practice of Infectious Diseases*, 886-913, Elsevier Saunders, Philadelphia; 2014.
4. Wang MC, Fan YH, Zhang YZ, Bregente CJB, Lin WH, Chen CA, et al. Characterization of uropathogenic *Escherichia coli* phylogroups associated with antimicrobial resistance, virulence factor distribution, and virulence-related phenotypes. *Infection, Genetics and Evolution*. 2023;114:105493.
5. Mishra S, Panda G. Molecular Characterization of Uropathogenic Markers in *Escherichia coli* Isolated from Pregnant Women having Urinary Tract Infection. *International Journal of Creative Research Thoughts*. 2022;10(2):540-554.
6. Fajfr M, Balik M, Cermakova E, Bostik P. Effective treatment for uncomplicated urinary tract infections with oral fosfomycin, single center four year retrospective study. *Antibiotics*. 2020;9(8):511.
7. Kowalska-Krochmal B, Mączyńska B, Rurańska-Smutnicka D, Secewicz A, Krochmal G, Bartelak M, et al. Assessment of the susceptibility of clinical Gram-negative and Gram-positive bacterial strains to fosfomycin and significance of this antibiotic in infection treatment. *Pathogens*. 2022;11(12):1441.
8. Ioannou P, Baliou S, Samonis G. Nanotechnology in the Diagnosis and Treatment of Antibiotic-Resistant Infections. *Antibiotics*. 2024;13(2):121.
9. Jin SE, Jin HE. Antimicrobial activity of zinc oxide nano/microparticles and their combinations against pathogenic microorganisms for biomedical applications: From physicochemical characteristics to pharmacological aspects. *Nanomaterials*. 2021;11(2):263.
10. Jabur NG. Isolation and identification of some bacterial species causing respiratory tract infections and evaluation their inhibitory efficiency using nanoprepared azithromycin and ciprofloxacin. M.Sc. thesis College of Science, University of Kerbala, Kerbala/ Iraq; 2021.
11. Bashi AM, Hussein MZ, Zainal Z, Tichit D. Synthesis and controlled release properties of 2, 4-dichlorophenoxy acetate-zinc layered hydroxide nano-hybrid. *Journal of Solid State Chemistry*. 2013;203:19-24.
12. Al-Khateeb MB, Al-Wakeel MI, Al-Ghanimi AA. Effect of nanoprepared levofloxacin in inhibiting of *Klebsilla pneumoniae* Isolated from respiratory tract infections.
13. Silverstein RM, Webster FX, Kiemle DJ. *Spectrometric Identification of Organic Compounds* [Internet]. 7 Ed. John Wiley and Sons. INC. Printed in United States of America; 2005.
14. Mir M, Leite FL, Herrmann Junior PSDP, Pissetti FL, Rossi AM, Moreira EL, et al. XRD, AFM, IR and TGA study of nanostructured hydroxyapatite. *Materials Research*. 2012;15:622-627.
15. Egorov NS. *Antibiotics: A scientific approach*. Mir Publishers; 1985.
16. Rahdar M, Rashki A, Miri HR, Ghalehnoo MR. Detection of pap, sfa, afa, foc, and fim

- adhesin-encoding operons in uropathogenic *Escherichia coli* isolates collected from patients with urinary tract infection. Jundishapur Journal of Microbiology. 2015;8(8).
17. Fardood ST, Ramazani A, Joo SW. Eco-friendly synthesis of magnesium oxide nanoparticles using arabic Gum. J. Appl. Chem. Res. 2018;12(1):8-15.
  18. Falagas ME, Vouloumanou EK, Samonis G, Vardakas KZ. Fosfomycin. Clinical Microbiology Reviews. 2016;29(2):321-347.
  19. Marino A, Stracquadanio S, Bellanca CM, Augello E, Ceccarelli M, Cantarella G, et al. Oral fosfomycin formulation in bacterial prostatitis: New role for an old molecule-brief literature review and clinical considerations. Infectious Disease Reports. 2022;14(4):621-634.
  20. Al-Fatlawi NG, AL-Ghanimi AA. Preparation and characterization of nano hybrid azithromycin and evaluate its inhibitory activity on locally isolated *Klebsiella pneumoniae*. Biochem Cell Arch. 2021;21(2):4775-4783.
  21. Kumar Nirmal, Das Satyajeet, Jyoti Anupam, Kaushik Sanket. Synergistic effect of silver nanoparticles with doxycycline against *Klebsiella pneumoniae*. Int J Pharm Pharm Sci. 2016;8(7):183-186.

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